Supplementary figure S1: EZH2 transient recruitment to active cis-regulatory elements. ChIP-seq data for the following regions chosen as example, Chr3: 68,257-74,244 (A) N1 and (B) N2 and Chr18: 59,908-61,868 (C) N1 and (D) N2 obtained before treatment and after 7 and 56 days on Ibrutinib. ChIP-seq has been performed for H3K4me3, H3K27me3 and EZH2, (A) and (C) H3K27ac patient R3 and (B) and (D) H3K27ac patient N3.

Supplementary figure S2: Global loss of H3K27ac and H3K27me3 after prolonged exposure to Ibrutinib. (A) Heat map for H3K27ac positive regions after up to 56 days on Ibrutinib. Peaks are ranked based on fold change (FC) at 56d versus 0d and separated based on FC = 1.5 (decreased at the bottom, increased at the top). Average peak intensities for the corresponding peak fractions are shown on the right part of the figure. (B) Quantification of immunoblot for H3K27me3 (Two RRs and 2 TNs) up to 12 months on Ibrutinib.

Supplementary figure S3: Promoters and non-promoters are occupied by different families of transcription factors and co-regulators. Cis-elements enriched for the H3K4me3 mark were screened for the presence of validated binding sites for transcription factors and co-regulators (ReMap database). For each protein of the database, data are expressed in total amount of sequences containing a validated binding site for the specified factor and sorted based on the data obtained for promoters (proteins with the highest number of associated sequences on the left). For example, the transcription factor SPI1 is found in 4322 promoters and 4552 non-promoters cis-elements enriched for H3K4me3 in CLL cells (R2).

Supplementary figure S4: Non-promoter sequences are enriched for DNA-binding motifs of B-cell specific transcription factors. Cis-elements enriched for the H3K4me3 mark were screened for the presence of validated binding sites for transcription factors and co-regulators (ReMap database). For each protein of the database, data were expressed as a score calculated using the highlighted equation allowing comparison between promoter and non-promoter sequences.

Supplementary figure S5: H3K4me3+/EZH2+ sequences are depleted from SMAD2-4-containing and bivalent cis-regulatory elements. (A) Comparison between peak scores obtained from REMI analysis separating EZH2 positive and negative sequences before treatment in N1 (equation (2) in supplemental methods). Peaks associated with (B) EZH2, (C) KDM5A and IRF3, (D) SMAD4 shown as a ratio to the total number of peaks (REMI analysis) in the 7

subgroups of peaks as analysed for N1. (E) Methodology to isolate H3K4me3-only peaks associated with SMAD proteins in N1 and N2. (F) *In vitro* analysis of CLL cells response to TGFB1 and WNT5A. Cells were treated for 4 hours with the indicated molecules and CXCR4 and RAC1 mRNA levels were measured.

Supplementary figure S6: Cis-elements containing validated SMAD binding sites are protected from EZH2 recruitment. (A) Gene ontology analysis of the peaks selected in Supplemental figure 5E. The red and blue squares correspond to the presence or absence of a gene, respectively. (B) REMI analysis comparing score for promoters versus CpG islands (equation (2) in supplemental methods). The bed file for promoters was generated in R with 1000 bp upstream and 500 bp downstream of transcription start site (TSS) for hg19 (UCSC). The CpG island bed file was also downloaded from UCSC (see Material and Methods).

Supplementary figure S7: Creation of a reference profile for H3K4me3. Score for N1 versus the average of N1-3, R1-2 patients before treatment. Cis-elements enriched for the H3K4me3 mark were screened for the presence of validated binding sites for transcription factors and coregulators (ReMap database). For each protein of the database, data were expressed as a score calculated using the highlighted equation. Details of the calculated scores are presented in supplementary table 5.

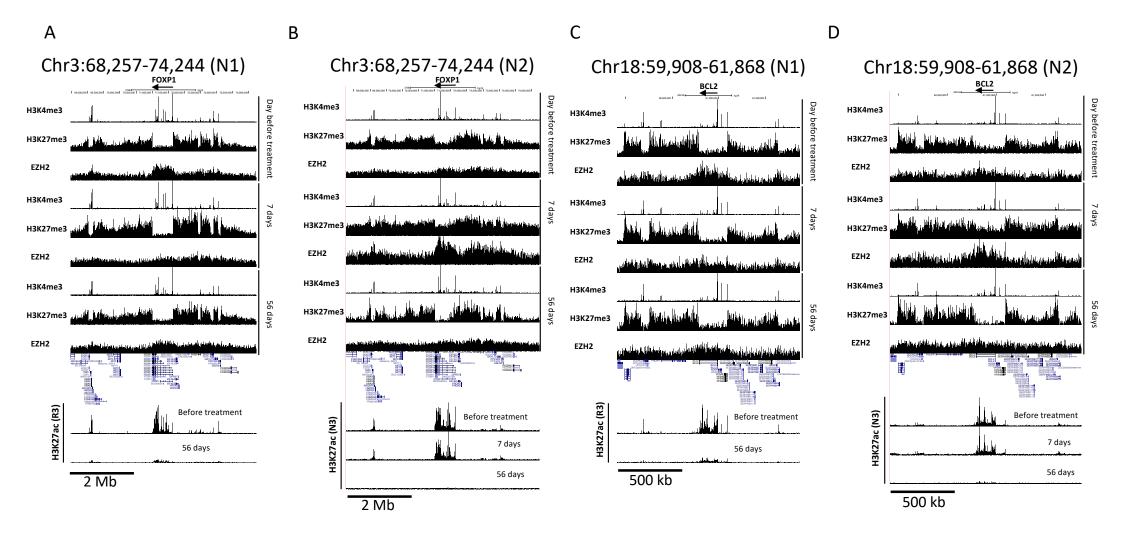
Supplementary figure S8: H3K4me3 positive cis-elements are differentially affected by Ibrutinib treatment. (A) promoters and (B) non-promoters heat maps for R1, N1, N2 and N3. Peaks were ranked based on fold change (FC) at 56d versus 0d and separated based on FC=1.5 (decreased at the bottom, increased at the top).

Supplementary figure S9: Subclasses of H3K4me3 positive elements are differentially affected by Ibrutinib treatment. (A) Promoters and non-promoters repartition of the H3K4me3 peaks for SMAD2,3 and 4 (group D). The ranked peaks were binned into 1000 peak subsets based on fold change and each subset analysed in REMI to assess the evolution of the number of peaks associated with the factors from SMAD subclass calculated as score (equation (3) in supplemental methods). (B) Heat maps including H3K4me3 at 0 and 56 days as well as H3K27me3 before treatment. The peaks are ranked based on H3K27me3 intensity. Regions positive for EZH2 (ReMap) are shown as barcodes on the right part of each heat map.

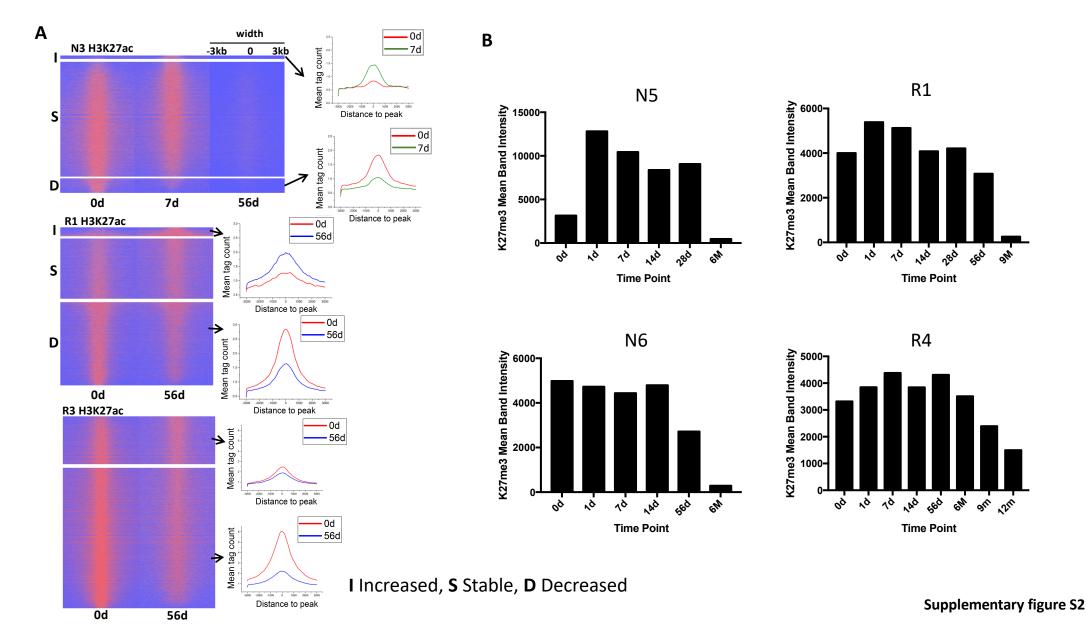
Supplementary figure S10: Evolution of H3K4me3 peaks number in CLL cells from patients the day before the start of Ibrutinib treatment (A) Variation in the number of H3K4me3 peaks associated with each factor of the ReMap database between N1-3 and R1-2 before treatment. The data were sorted from the most variable to the least variable. (B) Number of H3K4me3 peaks associated with EZH2 or SUZ12 for N1-3 and R1-2 before treatment.

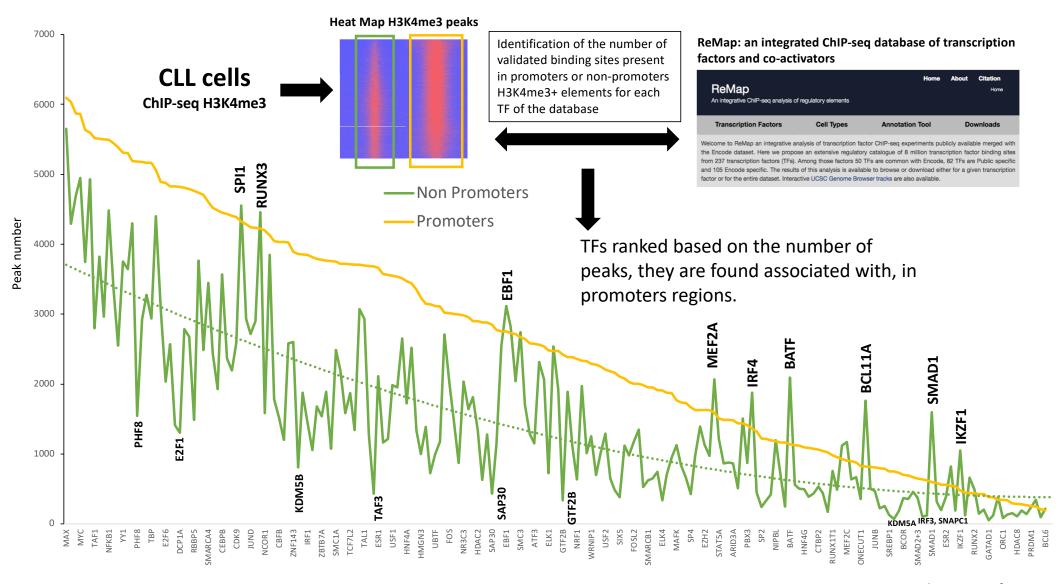
Supplementary figure S11: Identification of genes fully silenced in treatment-naïve patients only Gene ontology analysis for R1 and R2-only specific peaks, as described in figure 6A associated with EZH2 (annotated to the closest gene). The red and blue squares correspond to the presence and absence of a gene, respectively.

Supplementary figure S12: TIAM1 locus is silenced in treatment naive but not in relapsed/refractory patients before treatment. ChIP-seq data for H3K4me3 and H3K27me3 in TN and RR patients at the TIAM1 locus.



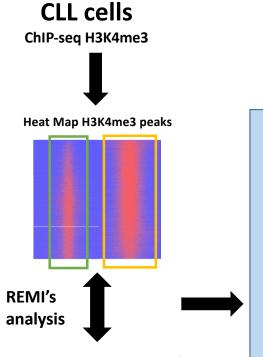
Supplementary figure S1





Supplementary figure S3

Calculation of a score to compare the variation in binding-factors composition of H3K4me3+ sequences between non-promoter and promoter regions



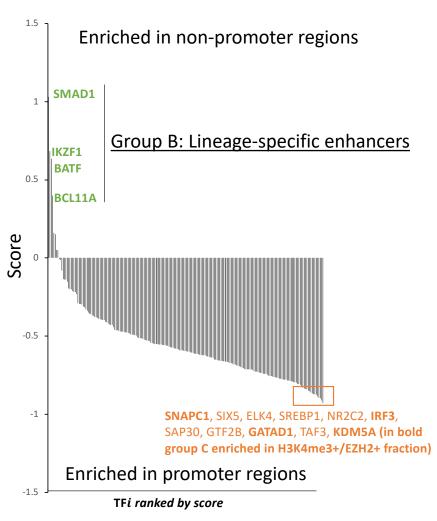
ReMap: an integrated ChIP-seq database of transcription factors and co-activators

An integrative ChiP-seq analysis of regulatory elements							
Transcription Factors	Cell Types	Annotation Tool	Downloads				
Welcome to ReMap an integrative and the Encode dataset. Here we propose from 237 transcription factors (TFs). A and 105 Encode specific. The results factor or for the entire dataset. Interact	e an extensive regulator mong those factors 50 of this analysis is availal	y catalogue of 8 million transcr TFs are common with Encode, ble to browse or download eithe	iption factor binding site 82 TFs are Public specifi				

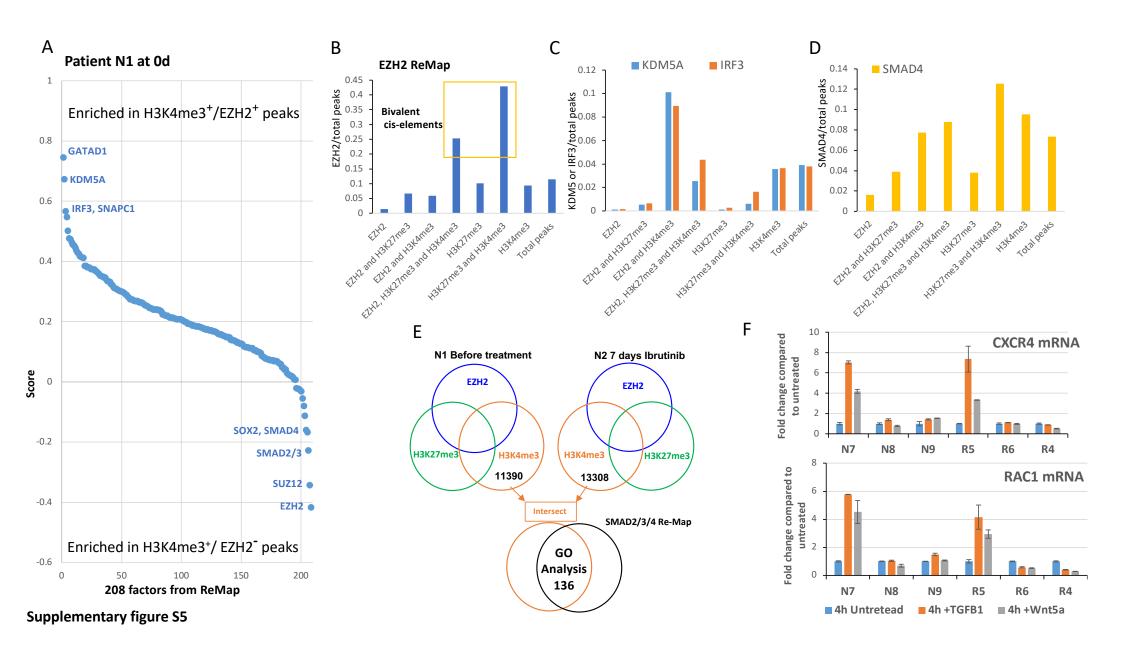
The **SCORE** for a certain TF can be described by the following equation:

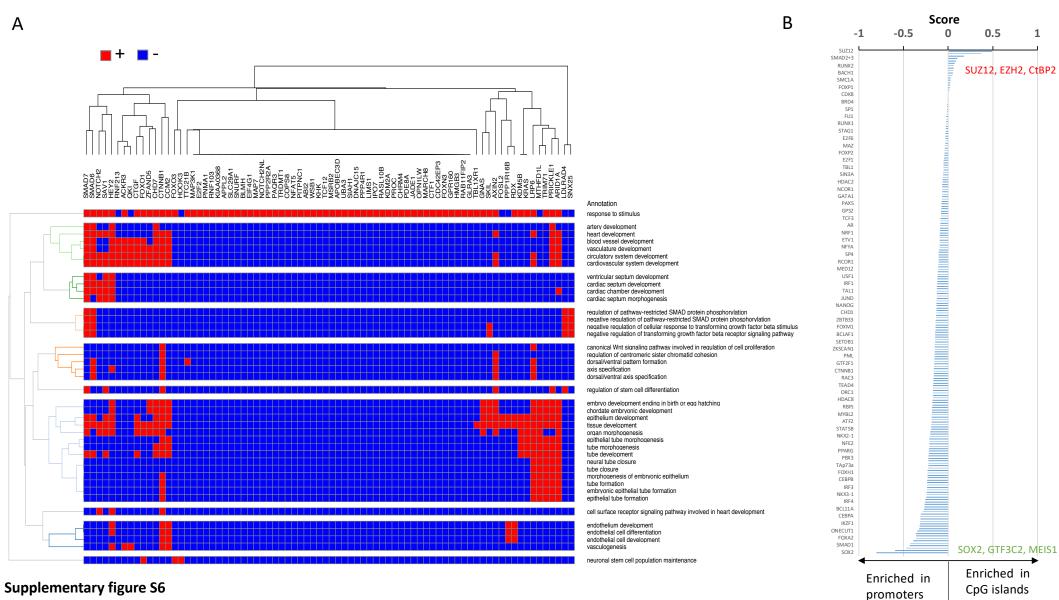
$$S_i = \frac{NP_i}{NP_t} / \frac{P_i}{P_t} - 1$$

where NP_t and P_t are the total number of validated-binding sites for the non-promoter and promoter groups, respectively. NP_i and P_i are the number of non-promoter and promoter sequences containing a validated-binding site for the TFi, respectively.



Supplementary figure \$4





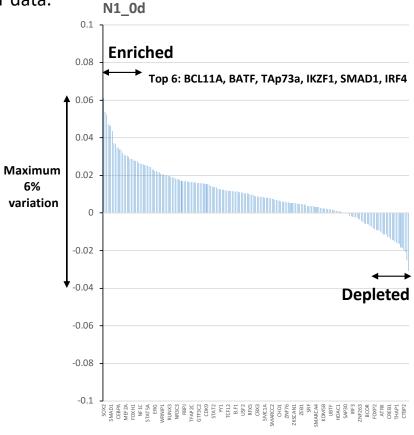
Data from N1 has been chosen because it represents the "middle point" when compared to the other data.

	R1	N2	R2	N3	N1	Average
ARID3A	3261	3276	3392	3689	3481	3419.8
ARNT	803	753	861	854	834	821
AR	11134	10900	11612	11732	11502	11376
ATF1	4099	4027	4162	4328	4179	4159
ATF2	6892	6523	7114	8128	7414	7214.2
ATF3	6573	6188	6750	6727	6557	6559
7015443	10747	0704	11224	11045	10772	1072
ZNF143	10747	9781	11324	11045	10773	10734
ZNF263	7566	6968	7876	7454	7434	7459.6
ZNF76	2764	2765	2969	2856	2859	2842.6
Total peaks	21174	18398	22025	23623	21836	21411.2

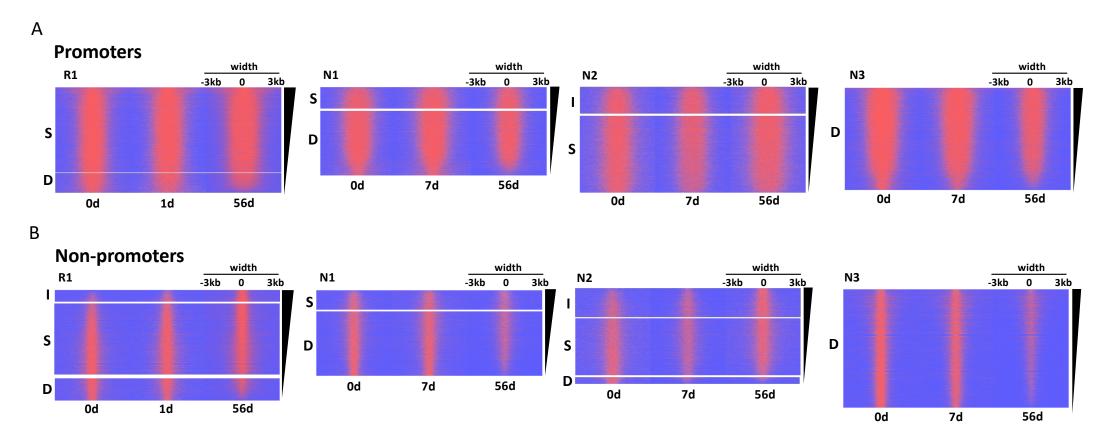
The **SCORE** for a certain TF can be described by the following equation:

$$S_i = \frac{N1_i}{N1_t} / \frac{A0_i}{A0_t} - 1$$

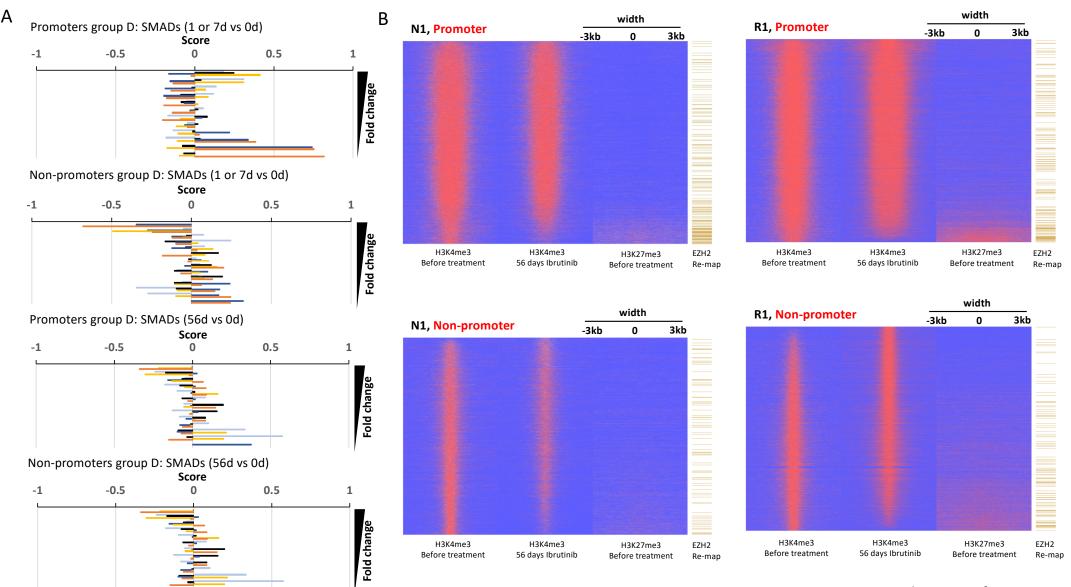
where $N1_t$ and $A0_t$ are the total number of validated-binding sites for the N1 peaks and average peak number for No Ibrutinib controls, respectively. $N1_i$ and $A0_i$ are the number of N1 peaks and average peak number for No Ibrutinib controls containing a validated-binding site for TFi, respectively.



REMI analysis

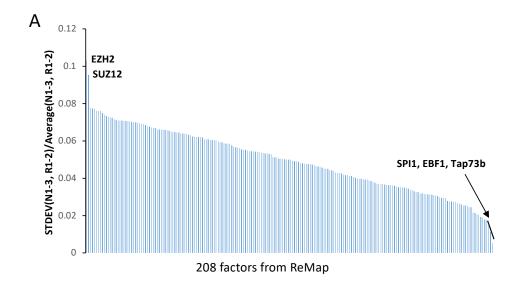


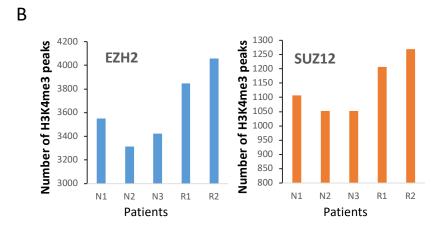
I Increased, **S** Stable, **D** Decreased



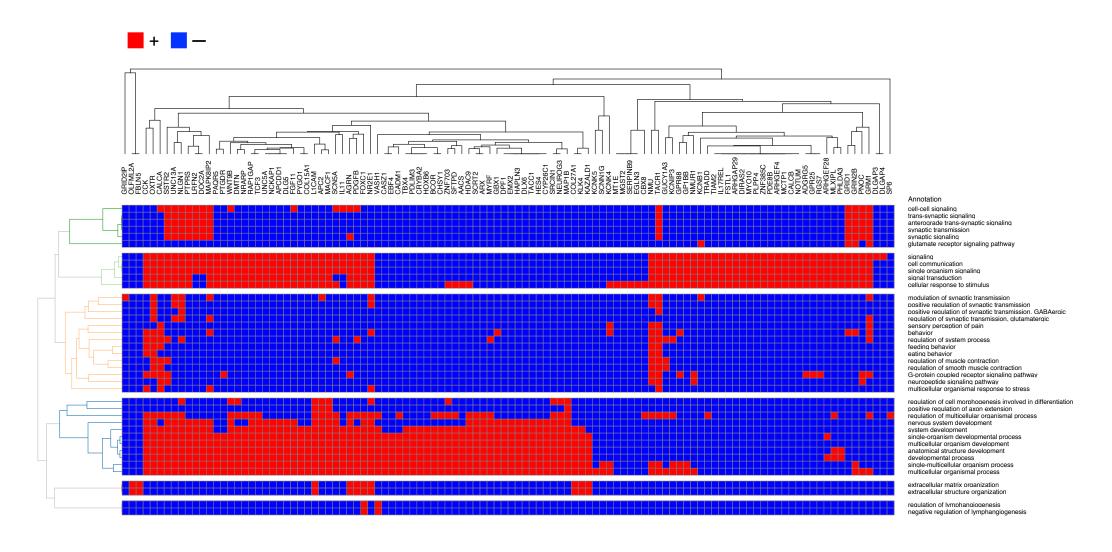
■N1 ■N2 ■N3 ■R1 ■R2

Supplementary figure S9





Supplementary figure S10



Supplementary figure S11

chr21:31,995,780-33,456,865

